

BMP1.3 protein as potential target in treatment of fibrosis

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ABSTRACT:

Bone morphogenetic protein 1 (BMP1) belongs to the procollagen C proteinase (PCP) family of proteases involved in development and pattern formation in various organisms. BMP1 proteinases mediate the cleavage of carboxyl peptides from procollagen molecules, which is a crucial step in fibrillar collagen synthesis. From six described alternatively spliced variants of human *Bmp1* gene, only BMP1.3 protein was detected in human plasma and elevated plasma levels of this protein were found in pathological conditions such as chronic kidney disease and acute myocardial infarction. Since BMP1 is required to convert pro-collagen to collagen, its inhibition is a potential intervention for treating fibrosis. Inhibition of BMP1.3 was shown to decrease the progression of liver fibrosis in an animal model of liver cirrhosis. One of the major inflammatory signaling molecules involved in fibrogenesis in various organs is transforming growth factor beta 1 (TGFβ1), which expression is elevated in various models of induced fibrosis. Many studies have revealed that BMP1 proteases play a key role in regulation of TGFβ activation. Here, we discuss BMP1.3 inhibition as a potential treatment in different pathological conditions related to the fibrosis. Testing BMP1.3 inhibition in these models indicates that the anti-BMP1.3 antibody targets relevant pathways in the development of fibrosis in different organs.

KEYWORDS: Bone morphogenetic protein 1 (BMP1), fibrosis, procollagen, transforming growth factor, chronic kidney disease

SAŽETAK:

BMP1.3 PROTEIN KAO POTENCIJALNA META U LIJEČENJU FIBROZE
 Koštani morfogenetski protein 1 (engl. BMP1) pripada obitelji C-proteinaza prokolagena (engl. PCP) uključenih u oblikovanje i razvitak različitih organizama. BMP1 proteinaze posreduju cijepanje peptida na karboksilnom kraju prokolagena, što je presudan korak u sintezi kolagena. Od šest opisanih varijanti humanog *Bmp1* gena, samo BMP1.3 protein cirkulira u humanoj plazmi, a povišene razine ovog proteina otkrivene su u patološkim stanjima kao što su kronična bolest bubrega i akutni infarkt miokarda. Budući da je BMP1 protein potreban za pretvorbu pro-kolagena u kolagen, njegova inhibicija predstavlja potencijalnu metodu u liječenju fibroze. Pokazano je da inhibicija BMP1.3 smanjuje napredovanje fibroze jetre u animalnom modelu ciroze jetre. Transformirajući čimbenik rasta beta 1 (engl. TGFβ1) jedna je od glavnih upalnih molekula uključenih u fibrogenezu različitih organa, a njegova je ekspresija povišena u različitim modelima inducirane fibroze. Mnoge studije pokazale su da BMP1 proteaze imaju ključnu ulogu u regulaciji aktivacije TGFβ. U ovom radu opisujemo inhibiciju BMP1.3 kao moguću terapiju različitih patoloških stanja povezanih s fibrozom. Istraživanje inhibicije BMP1.3 u tim modelima pokazuje da protutijela specifična za BMP1.3 djeluju na zajedničke puteve važne za razvoj fibroze u raznim organima.

KLJUČNE RIJEČI: koštani morfogenetski protein 1 (BMP1), fibroza, pro-kolagen, transformirajući čimbenik rasta, kronična bolest bubrega

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BMP1 ISOFORMS

Bone morphogenetic protein 1 (BMP1) was first isolated from bovine bone protein extract and appeared to be a regulatory molecule structurally and functionally different from other BMP proteins, which are mainly members of transforming growth factor (TGF) β protein family capable to induce cartilage formation *in vivo* (1). In contrast to the BMPs from the TGF β family, BMP1 is a protein belonging to the family of proteases implicated in development and pattern formation in various organisms, namely procollagen C proteinase (PCP) family (2). Phylogenetically, gene encoding BMP1 is closely related to the evolutionary old family of astacin-like genes described in the nematode worm *Caenorhabditis elegans*, where 40 genes encoding astacin-like proteins were discovered. From these 40 nematode-astacin (NAS) genes, gene *nas-39* is structurally identical to the human *Bmp1* gene, whose domain structure remained conserved during the evolution, probably even in the common ancestor of nematodes and vertebrates (3). The cDNA sequence of BMP1 showed high degree of similarity also to the *Drosophila* Tolloid metalloproteinase (4). In human and mice, the gene encoding BMP1 produces alternatively spliced transcripts with preserved domain structure, where the long isoform has an organization of domains identical to the *Drosophila* Tolloid and is designated mammalian Tolloid (mTLD)

(5). Further study (6) described six alternatively spliced variants of human *Bmp1* gene, named *Bmp1.1* do 1.6, where the *Bmp1.3* isoform is the longest and corresponds to mTLD, as well as to the *nas-39* from *C. elegans* (3, 7). Of all BMP1 isoforms, only BMP1.3 protein was found to circulate in humans and it was identified in human plasma samples by liquid chromatography-mass spectrometry (LC-MS) (8).

Structurally, BMP1 proteins are members of the astacin subgroup of metzincin metalloproteases, which contain a N-terminal prodomain followed by a catalytic astacin-like protease domain and one or more EGF (epidermal growth factor-like) and CUB (complement subcomponents C1r/C1s – embryonic sea urchin protein Uegf – BMP1) domains (5, 9). EGF and CUB domains are non-catalytic domains which promote protein-protein interactions. Studies on recombinant truncated forms of protein revealed that CUB1 domain is necessary for BMP1 secretion, and CUB2 domain, together with the protease domain, for C-proteinase activity (10, 11). Besides BMP1 and mTLD, there are two genetically distinct proteins named tolloid-like (TLL)-1 and TLL-2. Together with BMP1 and mTLD, they belong to the family of mammalian BMP1/TLD-like proteases which share common domain structure, but differ in C-terminal amino acid sequences (12) (Fig.1).

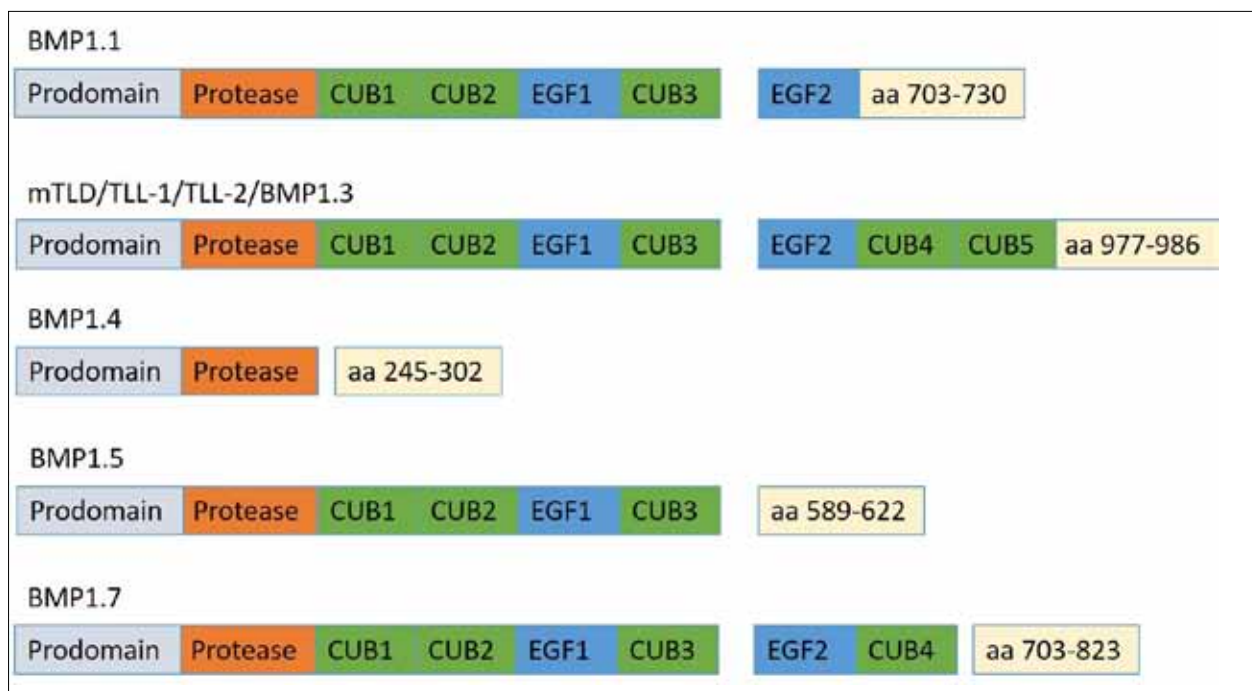


Figure 1. Schematic representation of BMP1 isoforms and their domain structure (adapted from (8))

BMP1/TLD-like proteases are involved in processing a wide range of extracellular matrix (ECM) precursors required for normal tissue assembly (13). Examples of such ECM precursor proteins are procollagens I-III, minor fibrillar procollagens, small leucine-rich proteoglycans (such as decorin, osteoglycin, biglycan), basement membrane components (laminin 332, collagen VII) and mineralization factors (dentin sialophosphoprotein, dentin matrix protein) (14). Further, BMP1/TLD-like proteases are engaged in release of TGF β superfamily members from their inhibitory complexes (e.g. TGF β 1, BMP2, -4 and -7), which in turn regulates developmental patterning and tissue homeostasis (13). In mice, *Bmp1* gene appears to be required for normal embryo development, and homozygous mutants with complete deletion of *Bmp1* are lethal because of herniation of the gut combined with failure of ventral body wall closure (15, 16). Early lethality of conventional *Bmp1* knockout mice required development of conditional knockouts with tissue-specific ablations of *Bmp1* expression in adult organism (16).

After discovery of BMP1.3 protein isoform circulating in human plasma, separated from its pro-domain (8), elevated levels of BMP1.3 have been found in pathological conditions such as chronic kidney disease (8) and acute myocardial infarction (17). Moreover, inhibition of BMP1.3 by specific antibodies reduced extent of experimentally induced liver fibrosis (18). BMP1.3 is also involved in bone fracture repair, where its inhibition delayed fracture healing (19). These findings provided the rationale for developing and testing the efficacy of BMP1.3 inhibition for treating fibrosis, a condition where therapy targeted to the BMP1.3 inhibition could reverse adverse effects due to the fibrotic changes in the particular organ. For this purpose, a monoclonal antibody was generated in mice immunized with a specific BMP1.3 synthetic peptide (C-terminal amino acids 972 to 986 unique for this isoform; RYTSTKFQDTLHSRK) (17, 18). Additionally, polyclonal antibody against mature BMP1.3 was generated in rabbits immunized with synthetic BMP1.3 peptide (amino acids 759 to 772; TSPNWPKYPSKKE) specific for mature domain of the protein (8). For monoclonal antibody production, the spleen from immunized Balb/c mice were used for *in vitro* hybridoma cell production. Cell culture supernatants from clones which produced the best monoclonal antibodies were collected and purified on Protein G affinity column. For polyclonal antibody production, New Zealand White rabbits were immunized with BMP1.3 peptide. Serum was collected between booster injections and antibody titer was determined. Appropriate aliquots of sera were affinity purified on Protein G column. These antibodies were used to explore the effects of BMP1 inhibition on liver fibrosis, chronic kidney disease, myocardial infarction and congenital muscle dystrophy in different animal models.

FIBROSIS AND THE ROLE OF TGF β

Fibrosis is defined as a reparative or reactive process characterized by the formation of excess of fibrous connective tissue, resulting in progressive remodelling of tissue or an organ (20). The process of wound healing after injury includes remodeling of extracellular matrix (ECM), where, in the early phase of regeneration, a provisional ECM is formed by cross-linking fibrin, fibronectin, fibrinogen and proteoglycans (21, 22). Although fibrogenic response is needed as a part of tissue repair process for restoring and maintaining an organ function after injury (scarring), an exacerbated fibrogenic response can shift this process to the chronic fibrosis which finally leads to destruction of normal organ architecture (21, 23).

Fibrosis is often triggered by an acute or chronic inflammatory process. Among major contributors to the induction of pathological fibrosis during chronic phase of inflammation are members of TGF β protein family (namely, TGF β 1, -2 and -3), whose suppression was sufficient to block experimentally induced fibrogenesis in various models. Among them, TGF β 1 is most important factor in tissue repair during inflammation and fibrosis (20). During the early phase of wound healing, TGF β 1 promotes collagen and fibronectin production and ECM formation; however, it acts also as a general suppressor of excessive inflammatory response, as seen from mice with inactivated *Tgfb1* gene (20, 24). Blockade of TGF β 1 production by antisense RNA attenuated experimentally induced liver fibrosis *in vitro* and *in vivo* (25). Similar effect was observed in a model overexpressing BMP7, which in turn negative influences *Tgfb1* expression via interference with several possible signalling mechanisms (26, 27). On the other hand, overexpression of *Tgfb1* in the liver lead to the development of hepatic fibrosis with multiple tissue lesions (28).

TGF β signalling pathways involve two types of receptors (TGFR1 and TGFR2) which act through so-called canonical (linked to intracellular Smad proteins) and non-canonical (non-Smad signalling molecules) pathways (29). For development of fibrosis, the most important is canonical signalling pathway which involves phosphorylation and activation of Smad2, Smad3 and Smad4 proteins (30), which in turn effect transcription of profibrotic genes (for example, α -smooth muscle actin (α -SMA), collagen I and tissue inhibitor of matrix deposition (*TIMP*)). Whereas Smad3 can bind directly to the Smad-binding elements within gene promoters, Smad2 and Smad4 need additional co-activators to act as regulators of gene transcription (31). TGF β activation is controlled by various inhibitory proteins, and for its regulation the crucial role play BMP1 proteases, which release active TGF β protein from the latent inhibitory complex (32). This complex consists of the TGF β linked to the prodomain (latency-associated peptide, LAP), which are together bound

to the latent TGFβ-binding protein (LTBP) and form the large latent complex (LLC) (33). BMP1 cleaves LTBP, enabling thus subsequent cleavage of LAP by matrix metalloproteinase (MMP) and activation of liberated TGFβ1 (32) (Fig. 2). A recent study using surface plasmon resonance assay demonstrated that CUB domain of BMP1 could be responsible for binding of TGFβ1,

increasing thus its signalling pathway (34). It has also been shown that BMP1 enhances TGFβ activity not only by proteolysis of its latent precursor, but also by cleavage of matricellular glycoprotein thrombospondin (TSP-1) (14). BMP1 is not the only activator of TGFβ, but is a significant factor in regulating its activity.

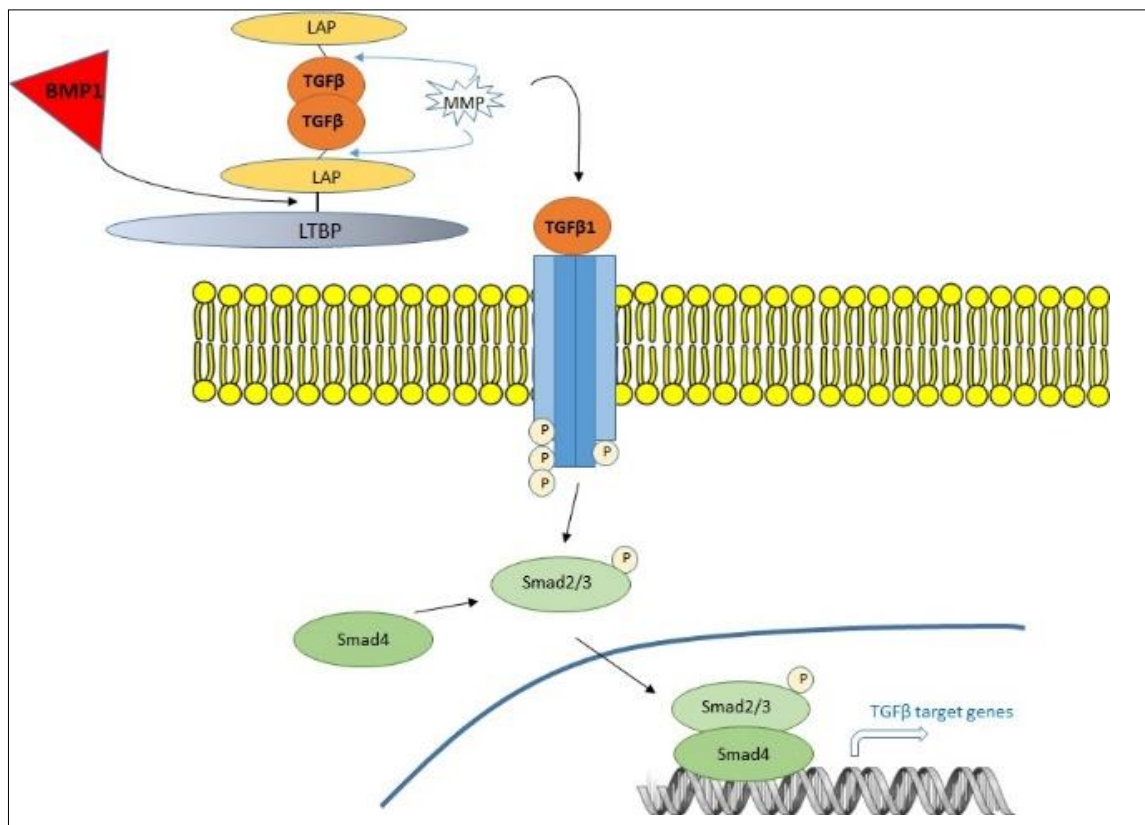


Figure 2. Activation of TGFβ and intracellular canonical signalling pathway. BMP1 cleaves latent TGFβ binding protein (LTBP), enabling thus subsequent cleavage of latency-associated peptide (LAP) by matrix metalloproteinase (MMP) and liberation and activation of TGFβ which then binds to receptor on cell membrane. Phosphorylation of TGFβ receptor upon binding of its ligand activates canonical Smad2/3 signalling pathway which activates transcription of corresponding genes.

LIVER FIBROSIS

Liver is among organs most frequently affected by fibrotic changes. Hepatic fibrosis (HF) usually results from an inflammatory process which first involves hepatocytes, but consequently leads to the activation of effector cells and excessive deposition of extracellular matrix. This process results finally in the liver cirrhosis, which can lead to the life-threatening complications (23). The most important role in development of liver fibrosis play hepatic stellate cells (HSCs), whose activation and proliferation during the liver injury leads to their transdifferentiation into myofibroblasts, which are characterized by expression of myogenic markers (α -SMA) (35) and increased ECM production (36). Activated HSCs are the major source of ECM in experimentally induced liver injury (37). Extensive studies on transgenic animal models revealed a number of key genes involved in liver fibrogenesis: genes regulating hepatocellular apoptosis/necrosis, genes regulating inflammatory response to the injury, genes regulating ROS generation, fibrogenic growth factors, vasoactive substances and adipokines (35). Among them, TGF β 1 has a crucial role in liver fibrogenesis in humans (38), stimulating transition of HSCs into myofibroblasts and inducing synthesis of ECM components (35) (Fig. 3). Quiescent HSCs are not the main source of TGF β 1; however, its

expression is significantly upregulated in these cells upon liver injury. Besides HSCs, there are additional cellular sources of TGF β 1 in liver tissue (hepatocytes, macrophages, platelets) (39). In activated HSCs, TGF β influences cytoskeletal organization and cellular migration through RhoA GTPase signalling (40), induces proliferative HSC response by a complex mechanism which involves PDGF β and PI3 kinase pathways, but also acts on hepatocytes by inducing expression of profibrogenic mediators (*PDGF*, *IL-15*, *TIMP-1*, *EGFR*) (41, 42).

The role of BMP1 in liver fibrosis is scarcely described in the literature. A recent paper published by our group (18) showed that inhibition of BMP1-3 protein by specific polyclonal antibodies reduced plasma levels of TGF β 1, suppressing thus its profibrotic effect and preventing progression of experimentally induced liver fibrosis *in vivo*. Further, treatment of human stellate cell line LX-2 (a model for HSCs *in vitro*) with BMP1.3 antibodies attenuated increase in *Col1* expression induced by TGF β treatment. In healthy liver, BMP1-3 is present mostly in sinusoidal epithelial cells, whereas in fibrotic liver it is found also in hepatocytes (18). This study suggested that BMP1.3 inhibition negatively influences release of TGF β 1 from its latent form, which in turn slows down the progress of fibrosis and could be considered as a new therapeutic approach.

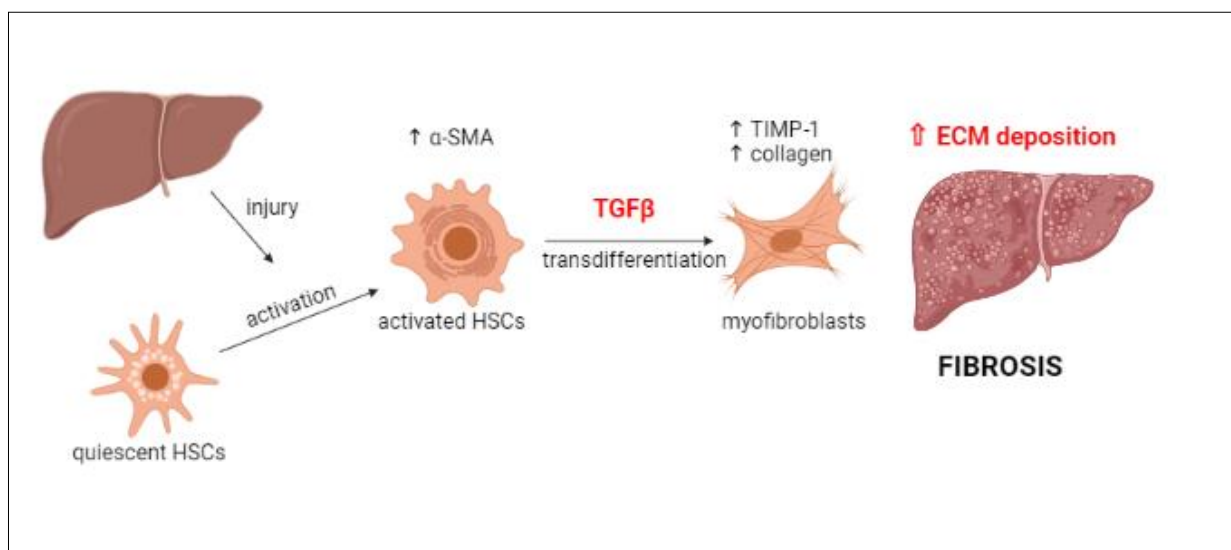


Figure 3. Schematic pathway of liver fibrosis development. Injury of liver parenchyma leads to the activation of quiescent hepatic stellate cells (HSCs), which, upon TGF β 1 signalling, transdifferentiate into myofibroblasts. Increased expression of TIMP-1 and collagen lead to the increased deposition of extracellular matrix (ECM) and development of liver fibrosis (image created with BioRender.com).

CHRONIC KIDNEY DISEASE

Chronic kidney disease (CKD) has a high prevalence (about 10% of worldwide population) and high mortality, usually developing progressively from chronic to end-stage renal disease, leading to kidney failure and finally requiring renal replacement therapies such as hemodialysis or kidney transplantation (43). CKD develops as a result of renal fibrotic process, characterized by increased ECM protein production and deposition of fibrotic matrix. Fibrosis affects all kidney compartments: glomeruli, tubulointerstitium and the vasculature (43), consequently resulting in the increased tissue stiffness and formation of scar tissue within the parenchyma, which ultimately leads to the kidney failure (44, 45).

Two most frequent diseases underlying renal fibrosis are hypertension and diabetes, but CKD often results from inflammatory kidney diseases, such as glomerulonephritis or inappropriate use of medications (46). As in other organs, the process of fibrosis is triggered by the chronic inflammatory process, characterized by the simultaneous tissue repair and remodelling (47). Excessive

accumulation of ECM proteins drives this process into fibrosis which disturbs normal organ function, interfering with normal regeneration of kidney structures. Similar to the fibrotic process in the liver described earlier, the main source of ECM in kidney are myofibroblasts. Myofibroblasts, which produce various types of ECM proteins (collagens, fibronectins, proteoglycans, etc.) can be derived from various cellular sources, but it seems that resident renal fibroblasts and hematopoietic cells which migrate to the kidney are their most important ancestors (48).

The initial trigger for renal fibrosis is infiltration of inflammatory cells which induce production of fibrogenic cytokines, such as IL-1, IL-8, TNF α , TGF β and other chemokines (49, 50). Among them, TGF β 1 is considered to have a primary role in induction and progression of CKD, similarly to its role in other organs. Besides its action on inducing ECM synthesis and reducing ECM degradation, TGF β 1 also promotes transition of endothelial cells and resident fibroblasts into myofibroblasts, which in turn promote further ECM deposition (31) (Fig. 4).

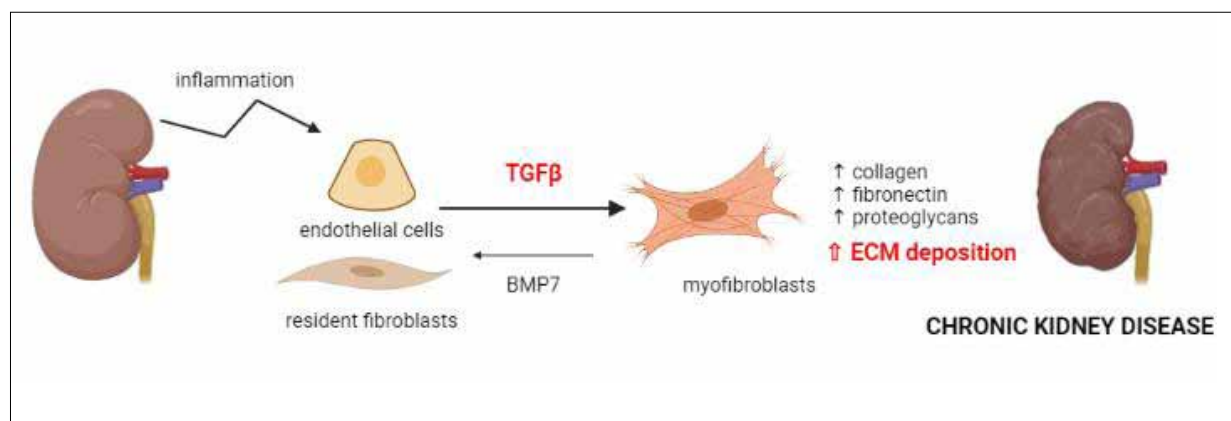


Figure 4. Schematic pathway of development of chronic kidney disease. Inflammatory process in kidney glomeruli and increased TGF β expression lead to the transdifferentiation of endothelial cells and resident fibroblasts into myofibroblasts, whereas BMP7 acts in the opposite direction. Myofibroblasts express increased amounts of collagen, fibronectin and proteoglycans, which together contribute to the increased ECM deposition and development of chronic kidney disease (image created with BioRender.com).

Numerous studies have shown increased expression of TGF β 1 in human kidney disease (51-53), whereas several animal studies have identified TGF β 1 as the predominant pathogenic factor in fibrogenesis regulation (31, 54-56). On the other hand, another mediator in kidneys, bone morphogenetic protein 7 (BMP7), which belongs to the TGF β superfamily of proteins, has an antifibrotic and anti-inflammatory effect and was found to prevent loss of kidney function in an animal model of acute renal failure (57). In kidneys, BMP7 is not only involved in regulation of fibrotic mechanisms, but also plays a role in kidney differentiation (58), acting via type II BMP receptors (59). Further studies revealed that BMP7 counteracts TGF β by reversing the TGF β -induced epithelial-to-mesenchymal transition and thus inhibits accumulation of myofibroblasts, which would otherwise lead to the development of fibrosis and CKD (60).

Although TGF β 1 can induce renal fibrosis via activation of different signalling pathways (29), the critical role in developing CKD has its canonical, Smad-dependent signalling pathway. Smad proteins mediate intracellular signal transduction which promotes deposition of ECM and inhibits its degradation, disturbing thus the balance between ECM aggregation and degradation and promoting the development of fibrosis (53, 55, 61). The primary downstream mediators of TGF β 1 are Smad2 and Smad3, which have opposite effect on development of kidney fibrosis. Phosphorylation of profibrotic Smad3 and suppression of antifibrotic Smad2 enhance fibroblast proliferation, differentiation of myofibroblasts and production of ECM in the kidney. In parallel, BMP7, which in kidney has an effect opposite to the TGF β , acts via Smad1/5/8 signalling pathway (62). Since there is currently no satisfactory strategy for treating CKD and preventing its progression to end-stage renal disease, new therapeutic approaches are needed. Considering the role of BMP1 in releasing an active form of TGF β 1, the main contributor to the fibrotic processes (31, 32), inhibition of BMP1 is potentially promising approach for treating kidney fibrosis. Recently, BMP1 inhibitor has been shown to block the accumulation of ECM components, collagens type I and III and fibronectin in both *in vitro* and *in vivo* models (63). Our group previously demonstrated that addition of specific polyclonal antibody raised against BMP1.3 (long) isoform attenuated fibrosis in experimentally induced CKD in rat model (8). Moreover, inhibition of BMP1.3 by specific antibodies increased expression of *Bmp7*, which could have an additional effect on reversal of fibrotic symptoms, regarding the role of BMP7 in kidney fibrosis, which is opposite to the TGF β (8). These findings support the BMP1 antagonism as a potentially new therapeutical strategy for renal fibrosis and CKD treatment.

HEART FAILURE

Acute myocardial infarction (AMI), which is the main cause of heart failure, affects more than 23 million people worldwide

(64). Myocardial infarction often results in significant fibrotic scarring of the heart with concomitant loss of cardiac function. Progressive loss of contractile function following a cardiac injury is a consequence of the poor regenerative capacity of cardiomyocytes and their replacement by a collagen-based fibrotic tissue. Although mammalian heart has a regenerative potential for a brief period after birth, in adult organism, this capacity is lost (65). Multiple attempts have been made to regenerate the heart using various strategies (66-69). Remodelling of the myocardium surrounding the site of injury, which includes thickening (hypertrophy) and stiffening (fibrosis) of the ventricular wall, eventually results in impaired cardiac function (70). Initially, after MI, dead cardiomyocytes are cleared by macrophages and are progressively replaced by reparative cells, mainly fibroblasts. Cardiac fibroblasts undergo three phenotypic changes: differentiation into myofibroblasts, proliferation and the production of extracellular matrix proteins (71). Among proteins produced by activated cardiac fibroblasts, BMP1 and lysyl oxidase (LOX) play a key role in collagen cross-linking (72, 73). It was demonstrated that BMP1 activates LOX precursor to mature active form which is responsible for the cross-linking of collagen. This process enables formation of mature, insoluble extracellular matrix less prone to degradation (72) (Fig. 5). Increased level and activity of LOX has been observed in the myocardium of patients with heart failure, which correlated with increased collagen cross-linking and collagen content (74).

As stated already in this review, members of TGF β family play a significant role in processes of cell differentiation and proliferation, in particular after tissue damage, when TGF β proteins are critical in wound healing and tissue regeneration. It is well-known that TGF β is highly expressed in neonatal and adult murine heart, localized in both cardiomyocytes and the ECM (75), where it is implicated to have a significant role in an early angiogenesis and development of cardiovascular structures (76). Expression of all three TGF β isoforms is significantly upregulated upon myocardial infarction in several animal models of AMI (77). In the myocardium, TGF β is present in its latent form, and for the activation of its signal transduction cascade, it must be released from the latent complex. The main factor in cleavage of this latent complex is BMP1, which acts as a metalloproteinase and releases TGF β from ECM, enabling thus its further activation through other enzymes (matrix metalloproteinases) (33). Besides TGF β activation, BMP1 also converts pro-collagen into collagen, one of main constituents of ECM and also fibrotic tissue. It has been shown that the endogenous expression of BMP1 was significantly upregulated at the same time point when type I and type III procollagen and TGF β were upregulated, which is consistent with the role of BMP1 as a key player in procollagen biosynthesis and maturation (78).

In infarcted heart, TGF β modulates immune reaction and cardiomyocyte survival and regulates regenerative processes in

the heart following infarction (79). Activation of fibroblasts by TGF β is an important part of this regenerative process, both by promoting transdifferentiation of fibroblasts into myofibroblasts and by stimulating ECM protein synthesis (80). Although this process is necessary for tissue regeneration, excessive myofibroblast activity and ECM deposition lead to the development of cardiac fibrosis, associated with increased stiffness and diastolic dysfunction, leading to heart failure (81).

Due to its role in regenerative and fibrotic processes, TGF β is potentially interesting therapeutic target in myocardial infarction (79). Inhibition of one of its main activators, BMP1, could present a new therapeutic approach in treatment of cardiac fibrosis. Both BMP1.1 and its long isoform BMP1.3 convert many of ECM precursors into mature functional proteins which mediate collagen crosslinking (82). Recently, our group has found increased levels of circulating BMP1.3 in plasma of patients with AMI, suggesting the possible role of this protein in cardiac fibrosis, whereas studies in animal models of AMI strongly suggest that BMP1.3 inhibition could have a therapeutic benefit. The newly developed anti-BMP1.3 monoclonal antibody was tested in animal model of myocardial infarction and found to reduce the expression of *Lox* in the scars of treated mice, as well as other crosslinking indicators (17).

CONGENITAL MUSCLE DYSTROPHY

The congenital muscular dystrophies (CMDs) are rare neuromuscular disorders, caused by allelic mutations in different genes and characterized by different pathologic features, primary affecting the skeletal muscle. CMDs vary in clinical features, but common are dystrophic features found by muscle biopsy, including variations in muscle fiber size, fiber degeneration and increased fibrosis (83). Chronic inflammatory processes in dystrophic

muscle result in excessive accumulation of ECM components, leading to the replacement of muscle with fibrotic tissue (84). CMDs are classified in several groups, depending on source of protein defect. The most common form of CMD is laminin- α 2 chain-deficient muscular dystrophy (LAMA2 MD), also called merosin-deficient CMD (MDC1A), which accounts for about 30% of all CMD patients in Europe (83, 85). Early-onset MDC1A is an autosomal recessive disorder caused by mutation in *LAMA2* gene which leads to loss of α 2 subunit of laminin-211 protein. Laminins are heterotrimeric proteins composed of α , β and γ subunits, and laminin-211 (also called merosin) is predominantly expressed in skeletal muscle as an important tissue component of ECM (83, 86). Laminins are essential for basement membrane assembly and insufficient assembly of laminin network causes poor connection to the muscle fibers, which may be one of causes of MDC1A (87). Structural and functional integrity of basement membrane is crucial for regulation of cell interactions and processes which regulate differentiation of myofibroblasts, which are in turn critical for development of fibrosis (88).

Laminin-211 is expressed primarily in basement membranes of skeletal muscle and Schwann cells, but also in other tissues (heart, kidney, lung, stomach, placenta, testis) (89). Muscles lacking laminin-211 have signs of chronic inflammation and widespread fibrosis in the interstitial space (89), and there is substantially high expression of ECM genes in CMD, regardless of histological changes, supporting an early fibrosis in this disease (90). As the most significant driver of fibrosis appears TGF β , whose activation depends on proteolysis of latent inhibitory complex by BMP1 (32), but also on activation of integrins (89). Indeed, TGF β signalling increases early in CMD, stimulating fibroblasts to produce ECM components such as collagen and fibronectin

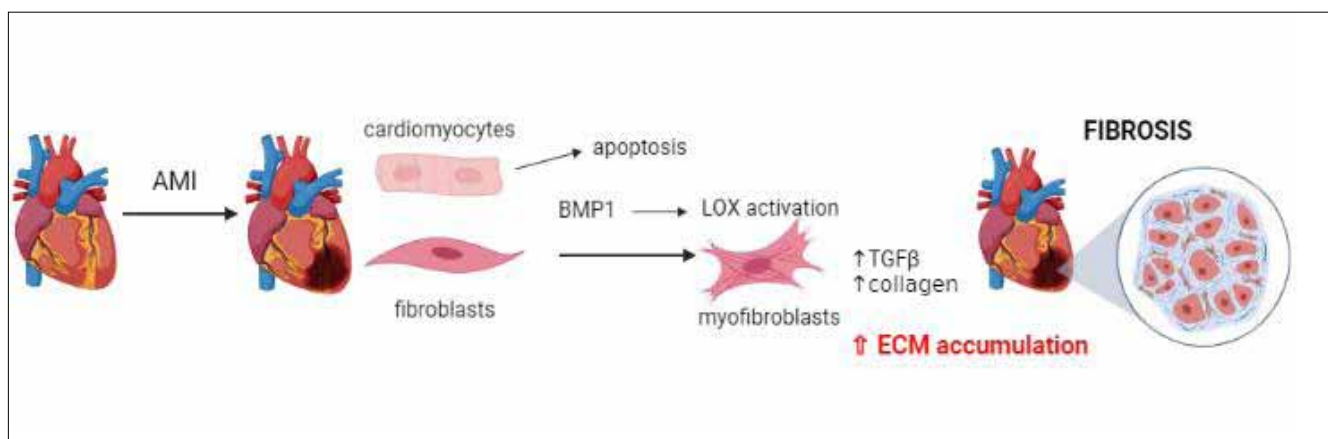


Figure 5. Schematic pathway of heart fibrosis development. Upon acute myocardial infarction (AMI), cardiomyocytes undergo apoptosis, and cardiac fibroblasts undergo transdifferentiation into myofibroblasts. Simultaneously, BMP1 activates lysyl oxidase (LOX) precursor into mature form, which is responsible for collagen cross-linking. Increased expression of TGF β and collagen lead to the increased ECM accumulation and development of fibrosis (image created with BioRender.com).

and influencing negatively production of enzymes which degrade ECM (84, 91). TGF β signalling pathway is stimulated through increased activity of phosphorylated of Smad2/3, as seen in mouse model for MDC1A (91). Chronic dysregulation of TGF β signalling leads also to the transdifferentiation of myoblasts into myofibroblasts (Fig. 6). TGF β 1 seems to be a key player in this process through complex signalling mechanisms involving integrins (92) and sphingosine kinase pathway (93), along with increased expression of muscle fibroblast marker Tcf4 (94). Most therapeutical strategies for MDC1A are targeting TGF β 1 signalling pathway in order to alleviate fibrotic processes, for example, losartan, which is originally approved for treatment of hypertension as an AT $_1$ R blocker (89). Losartan (or its derivatives) acts on angiotensin-renin system, which indirectly inhibits TGF β 1 signalling and leads to amelioration of fibrosis, as demonstrated in mouse models of MDC1A (95, 96). However, although treatment with losartan reduced inflammation and fibrosis, it did not increase muscle weight and could not be considered as a single mode therapy for this disease but should be supplemented with growth-inducing therapy (89, 97). Other therapeutical strategies which are currently explored include protein replacement therapy with addition of recombinant laminin-111 (98), use of linker proteins, targeting of intracellular regulatory systems or genetic approaches (for review, see (85)). Another therapeutical approach could also be inhibition

of BMP1.3 by the use of specific antibodies, which already demonstrated its efficacy in reducing fibrosis in kidney (8) and liver (18). These studies are ongoing on animal models, by using DyW mice which are homozygous for the mutation in the laminin- α 2 gene and are widely used as an animal model for MDC1A (99, 100). Initial studies on DyW mice suggest that treatment with anti-BMP1.3 antibodies could improve DyW mice mobility, quality of life and life expectancy in comparison with mice without therapy (unpublished data).

CONCLUDING REMARKS

As proteolytic activators of TGF β , one of main mediators of fibrotic processes, a group of BMP1 proteins has an universal function in mammalian organism. Among multiple BMP1 isoforms, only BMP1.3 (the long isoform) has been found to circulate in human blood. In search for appropriate targets for treating fibrosis, inhibition of BMP1.3 via synthetic inhibitors or specific antibodies could be a promising solution. Results on animal models of liver fibrosis, chronic kidney disease and myocardial infarction suggest that anti-BMP1.3 antibodies could alleviate fibrotic symptoms in these indications, and also to slow the progression of congenital muscular dystrophy (Fig. 7). More studies on animal models are needed, as well as clinical studies in humans, which would encourage development of specific biological therapies for these diseases.

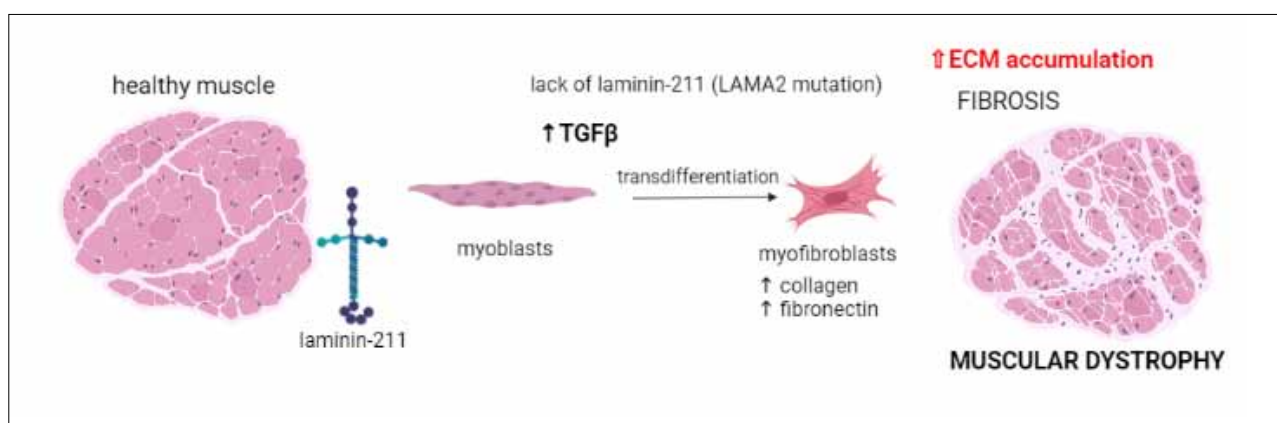


Figure 6. Development of muscular dystrophy. LAMA2 mutation causes lack of laminin-211, a protein crucial for proper basement formation in healthy muscle. Increased TGF β signalling in muscles lacking laminin-211 stimulate transdifferentiation of myoblasts into myofibroblasts which produce increased amounts of collagen and fibronectin and contribute to the ECM accumulation and development of fibrosis. Muscle fibers degenerate and muscular dystrophy develops (image created with BioRender.com).

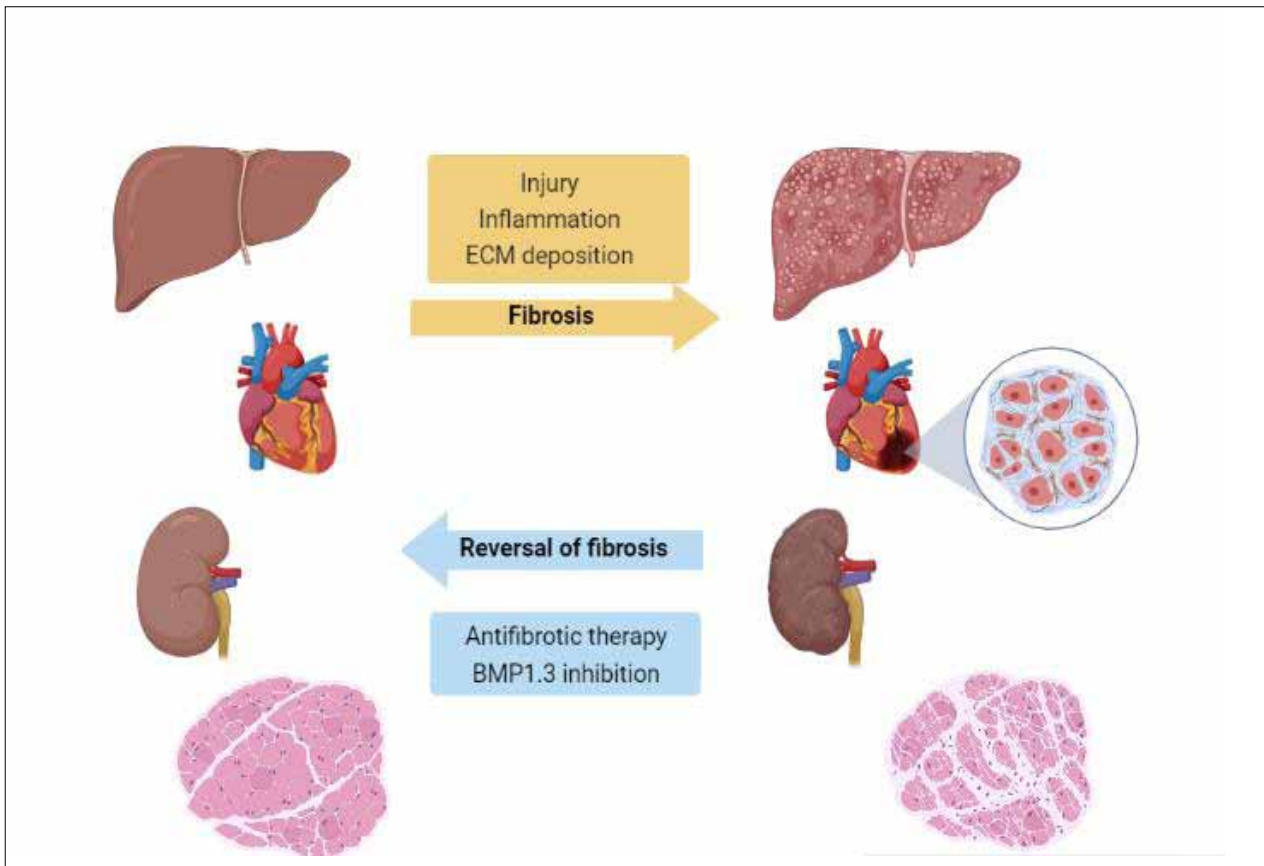


Figure 7. Examples of organs most frequently affected by chronic fibrosis. Injury and chronic inflammation accompanied by increased ECM deposition result in development of fibrosis, which finally lead to the loss of the function of particular organ (liver, heart, kidney, skeletal muscle). Antifibrotic therapy could relieve fibrotic symptoms and enable reversal of fibrosis. Among possible therapeutic approaches, inhibition of BMP1.3, which is one of the main activators of signalling cascade leading to development of fibrosis, appears to be a promising solution (image created with BioRender.com).

REFERENCES:

1. Wozney JM, Rosen V, Celeste AJ, Mitscock LM, Whitters MJ, Kriz RW, et al. Novel regulators of bone formation: molecular clones and activities. *Science*. 1988; 242:1528-34.
2. Kessler E, Takahara K, Biniaminov L, Brusel M, Greenspan DS. Bone morphogenetic protein-1: the type I procollagen C-proteinase. *Science*. 1996; 271:360-2.
3. Mohrlen F, Hutter H, Zwilling R. The astacin protein family in *Caenorhabditis elegans*. *Eur J Biochem*. 2003; 270:4909-20.
4. Shimell MJ, Ferguson EL, Childs SR, O'Connor MB. The *Drosophila* dorsal-ventral patterning gene *tolloid* is related to human bone morphogenetic protein 1. *Cell*. 1991; 67:469-81.
5. Takahara K, Lyons GE, Greenspan DS. Bone morphogenetic protein-1 and a mammalian tolloid homologue (mTld) are encoded by alternatively spliced transcripts which are differentially expressed in some tissues. *J Biol Chem*. 1994; 269:32572-8.
6. Janitz M, Heiser V, Bottcher U, Landt O, Lauster R. Three alternatively spliced variants of the gene coding for the human bone morphogenetic protein-1. *J Mol Med (Berl)*. 1998; 76:141-6.
7. Park JO, Pan J, Mohrlen F, Schupp MO, Johnsen R, Baillie DL, et al. Characterization of the astacin family of metalloproteases in *C. elegans*. *BMC Dev Biol*. 2010; 10:14.
8. Grgurevic L, Macek B, Healy DR, Brault AL, Erjavec I, Cipicic A, et al. Circulating bone morphogenetic protein 1-3 isoform increases renal fibrosis. *J Am Soc Nephrol*. 2011; 22:681-92.
9. Bond JS, Beynon RJ. The astacin family of metalloendopeptidases. *Protein Sci*. 1995; 4:1247-61.
10. Hartigan N, Garrigue-Antar L, Kadler KE. Bone morphogenetic protein-1 (BMP-1). Identification of the minimal domain structure for procollagen C-proteinase activity. *J Biol Chem*. 2003; 278:18045-9.
11. Petropoulou V, Garrigue-Antar L, Kadler KE. Identification of the minimal domain structure of bone morphogenetic protein-1 (BMP-1) for chordinase activity: chordinase activity is not enhanced by procollagen C-proteinase enhancer-1 (PCPE-1). *J Biol Chem*. 2005; 280:22616-23.
12. Scott IC, Blitz IL, Pappano WN, Imamura Y, Clark TG, Steigltz BM, et al. Mammalian BMP-1/Tolloid-related metalloproteinases, including novel family member mammalian Tolloid-like 2, have differential enzymatic activities and distributions of expression relevant to patterning and skeletogenesis. *Dev Biol*. 1999; 213:283-300.
13. Troilo H, Bayley CP, Barrett AL, Lockhart-Cairns MP, Jowitt TA, Baldock C. Mammalian tolloid proteinases: role in growth factor signalling. *FEBS Lett*. 2016; 590:2398-407.
14. Anastasi C, Rousselle P, Talantikite M, Tessier A, Cluzel C, Bachmann A, et al. BMP-1 disrupts cell adhesion and enhances TGF-beta activation through cleavage of the extracellular matrix protein thrombospondin-1. *Sci Signal*. 2020; 13.
15. Suzuki N, Labosky PA, Furuta Y, Hargett L, Dunn R, Fogo AB, et al. Failure of ventral body wall closure in mouse embryos lacking a procollagen C-proteinase encoded by *Bmp1*, a mammalian gene related to *Drosophila* *tolloid*. *Development*. 1996; 122:3587-95.
16. Ge G, Greenspan DS. Developmental roles of the BMP1/TLD metalloproteinases. *Birth Defects Res C Embryo Today*. 2006; 78:47-68.
17. Vukicevic S, Colliva A, Kufner V, Martinelli V, Moimas S, Vodret S, et al. Bone Morphogenetic Protein 1.3 inhibition supports cardiomyocyte survival and decreases scar formation after myocardial infarction *Nat Commun*. 2021; in press.
18. Grgurevic L, Erjavec I, Grgurevic I, Dumic-Cule I, Brkljacic J, Verbanac D, et al. Systemic inhibition of BMP1-3 decreases progression of CCl4-induced liver fibrosis in rats. *Growth Factors*. 2017; 35:201-15.
19. Grgurevic L, Macek B, Mercep M, Jelic M, Smoljanovic T, Erjavec I, et al. Bone morphogenetic protein (BMP)1-3 enhances bone repair. *Biochem Biophys Res Commun*. 2011; 408:25-31.
20. Weiskirchen R, Weiskirchen S, Tacke F. Organ and tissue fibrosis: Molecular signals, cellular mechanisms and translational implications. *Mol Aspects Med*. 2019; 65:2-15.
21. Pakshir P, Hinz B. The big five in fibrosis: Macrophages, myofibroblasts, matrix, mechanics, and miscommunication. *Matrix Biol*. 2018; 68-69:81-93.
22. Wight TN. Provisional matrix: A role for versican and hyaluronan. *Matrix Biol*. 2017; 60-61:38-56.
23. Rockey DC, Bell PD, Hill JA. Fibrosis--a common pathway to organ injury and failure. *N Engl J Med*. 2015; 372:1138-49.
24. Shull MM, Ormsby I, Kier AB, Pawlowski S, Diebold RJ, Yin M, et al. Targeted disruption of the mouse transforming growth factor-beta 1 gene results in multifocal inflammatory disease. *Nature*. 1992; 359:693-9.
25. Arias M, Sauer-Lehnen S, Treptau J, Janoschek N, Theuerkauf I, Buettner R, et al. Adenoviral expression of a transforming growth factor-beta1 antisense mRNA is effective in preventing liver fibrosis in bile-duct ligated rats. *BMC Gastroenterol*. 2003; 3:29.
26. Kinoshita K, Iimuro Y, Otogawa K, Saika S, Inagaki Y, Nakajima Y, et al. Adenovirus-mediated expression of BMP-7 suppresses the development of liver fibrosis in rats. *Gut*. 2007; 56:706-14.

27. Wang S, Hirschberg R. Bone morphogenetic protein-7 signals opposing transforming growth factor beta in mesangial cells. *J Biol Chem.* 2004; 279:23200-6.
28. Sanderson N, Factor V, Nagy P, Kopp J, Kondaiah P, Wakefield L, et al. Hepatic expression of mature transforming growth factor beta 1 in transgenic mice results in multiple tissue lesions. *Proc Natl Acad Sci U S A.* 1995; 92:2572-6.
29. Moustakas A, Heldin CH. Non-Smad TGF-beta signals. *J Cell Sci.* 2005; 118:3573-84.
30. Shi Y, Massague J. Mechanisms of TGF-beta signaling from cell membrane to the nucleus. *Cell.* 2003; 113:685-700.
31. Meng XM, Nikolic-Paterson DJ, Lan HY. TGF-beta: the master regulator of fibrosis. *Nat Rev Nephrol.* 2016; 12:325-38.
32. Ge G, Greenspan DS. BMP1 controls TGFbeta1 activation via cleavage of latent TGFbeta-binding protein. *J Cell Biol.* 2006; 175:111-20.
33. Robertson IB, Rifkin DB. Regulation of the Bioavailability of TGF-beta and TGF-beta-Related Proteins. *Cold Spring Harb Perspect Biol.* 2016; 8.
34. Predes D, Cruz JVR, Abreu JG, Mendes FA. CUB domain-containing protein 1 (CDCP1) binds transforming growth factor beta family members and increase TGF-beta1 signaling pathway. *Exp Cell Res.* 2019; 383:111499.
35. Bataller R, Brenner DA. Liver fibrosis. *J Clin Invest.* 2005; 115:209-18.
36. Puche JE, Saiman Y, Friedman SL. Hepatic stellate cells and liver fibrosis. *Compr Physiol.* 2013; 3:1473-92.
37. Tsuchida T, Friedman SL. Mechanisms of hepatic stellate cell activation. *Nat Rev Gastroenterol Hepatol.* 2017; 14:397-411.
38. Gressner AM, Weiskirchen R, Breitkopf K, Dooley S. Roles of TGF-beta in hepatic fibrosis. *Front Biosci.* 2002; 7:d793-807.
39. Dewidar B, Meyer C, Dooley S, Meindl-Beinker AN. TGF-beta in Hepatic Stellate Cell Activation and Liver Fibrogenesis-Updated 2019. *Cells.* 2019; 8.
40. Li L, Wang JY, Yang CQ, Jiang W. Effect of RhoA on transforming growth factor beta1-induced rat hepatic stellate cell migration. *Liver Int.* 2012; 32:1093-102.
41. Dooley S, Hamzavi J, Ciuculan L, Godoy P, Ilkavets I, Ehner S, et al. Hepatocyte-specific Smad7 expression attenuates TGF-beta-mediated fibrogenesis and protects against liver damage. *Gastroenterology.* 2008; 135:642-59.
42. Fabregat I, Moreno-Caceres J, Sanchez A, Dooley S, Dewidar B, Giannelli G, et al. TGF-beta signalling and liver disease. *FEBS J.* 2016; 283:2219-32.
43. Djudjaj S, Boor P. Cellular and molecular mechanisms of kidney fibrosis. *Mol Aspects Med.* 2019; 65:16-36.
44. Chung JY, Chan MK, Li JS, Chan AS, Tang PC, Leung KT, et al. TGF-beta Signaling: From Tissue Fibrosis to Tumor Microenvironment. *Int J Mol Sci.* 2021; 22.
45. Humphreys BD. Mechanisms of Renal Fibrosis. *Annu Rev Physiol.* 2018; 80:309-26.
46. Romagnani P, Remuzzi G, Glassock R, Levin A, Jager KJ, Tonelli M, et al. Chronic kidney disease. *Nat Rev Dis Primers.* 2017; 3:17088.
47. Wynn TA. Cellular and molecular mechanisms of fibrosis. *J Pathol.* 2008; 214:199-210.
48. Mack M, Yanagita M. Origin of myofibroblasts and cellular events triggering fibrosis. *Kidney Int.* 2015; 87:297-307.
49. Feghali CA, Wright TM. Cytokines in acute and chronic inflammation. *Front Biosci.* 1997; 2:d12-26.
50. Liu Y. Cellular and molecular mechanisms of renal fibrosis. *Nat Rev Nephrol.* 2011; 7:684-96.
51. Sharma K, Ziyadeh FN, Alzahabi B, McGowan TA, Kapoor S, Kurnik BR, et al. Increased renal production of transforming growth factor-beta1 in patients with type II diabetes. *Diabetes.* 1997; 46:854-9.
52. Yamamoto T, Nakamura T, Noble NA, Ruoslahti E, Border WA. Expression of transforming growth factor beta is elevated in human and experimental diabetic nephropathy. *Proc Natl Acad Sci U S A.* 1993; 90:1814-8.
53. Yoshioka K, Takemura T, Murakami K, Okada M, Hino S, Miyamoto H, et al. Transforming growth factor-beta protein and mRNA in glomeruli in normal and diseased human kidneys. *Lab Invest.* 1993; 68:154-63.
54. Border WA, Okuda S, Languino LR, Sporn MB, Ruoslahti E. Suppression of experimental glomerulonephritis by antiserum against transforming growth factor beta 1. *Nature.* 1990; 346:371-4.
55. Moon JA, Kim HT, Cho IS, Sheen YY, Kim DK. IN-1130, a novel transforming growth factor-beta type I receptor kinase (ALK5) inhibitor, suppresses renal fibrosis in obstructive nephropathy. *Kidney Int.* 2006; 70:1234-43.
56. Russo LM, del Re E, Brown D, Lin HY. Evidence for a role of transforming growth factor (TGF)-beta1 in the induction of postglomerular albuminuria in diabetic nephropathy: amelioration by soluble TGF-beta type II receptor. *Diabetes.* 2007; 56:380-8.
57. Vukicevic S, Basic V, Rogic D, Basic N, Shih MS, Shepard A, et al. Osteogenic protein-1 (bone morphogenetic protein-7) reduces severity of injury after ischemic acute renal failure in rat. *J Clin Invest.* 1998; 102:202-14.
58. Vukicevic S, Kopp JB, Luyten FP, Sampath TK. Induction of nephrogenic mesenchyme by osteogenic protein 1 (bone morphogenetic protein 7). *Proc Natl Acad Sci U S A.* 1996; 93:9021-6.

59. Bosukonda D, Shih MS, Sampath KT, Vukicevic S. Characterization of receptors for osteogenic protein-1/bone morphogenetic protein-7 (OP-1/BMP-7) in rat kidneys. *Kidney Int.* 2000; 58:1902-11.
60. Zeisberg M, Hanai J, Sugimoto H, Mammoto T, Charytan D, Strutz F, et al. BMP-7 counteracts TGF-beta1-induced epithelial-to-mesenchymal transition and reverses chronic renal injury. *Nat Med.* 2003; 9:964-8.
61. Bottinger EP, Bitzer M. TGF-beta signaling in renal disease. *J Am Soc Nephrol.* 2002; 13:2600-10.
62. Meng XM, Chung AC, Lan HY. Role of the TGF-beta/BMP-7/Smad pathways in renal diseases. *Clin Sci (Lond).* 2013; 124:243-54.
63. Bai M, Lei J, Wang S, Ding D, Yu X, Guo Y, et al. BMP1 inhibitor UK383,367 attenuates renal fibrosis and inflammation in CKD. *Am J Physiol Renal Physiol.* 2019; 317:F1430-F8.
64. Bui AL, Horwich TB, Fonarow GC. Epidemiology and risk profile of heart failure. *Nat Rev Cardiol.* 2011; 8:30-41.
65. Porrello ER, Mahmoud AI, Simpson E, Hill JA, Richardson JA, Olson EN, et al. Transient regenerative potential of the neonatal mouse heart. *Science.* 2011; 331:1078-80.
66. Bortolotti F, Ruozi G, Falcione A, Doimo S, Dal Ferro M, Lesizza P, et al. In Vivo Functional Selection Identifies Cardiotrophin-1 as a Cardiac Engraftment Factor for Mesenchymal Stromal Cells. *Circulation.* 2017; 136:1509-24.
67. Gabisonia K, Prosdocimo G, Aquaro GD, Carlucci L, Zentilin L, Secco I, et al. MicroRNA therapy stimulates uncontrolled cardiac repair after myocardial infarction in pigs. *Nature.* 2019; 569:418-22.
68. Nakada Y, Canseco DC, Thet S, Abdisalaam S, Asaithamby A, Santos CX, et al. Hypoxia induces heart regeneration in adult mice. *Nature.* 2017; 541:222-7.
69. Thavapalachandran S, Grieve SM, Hume RD, Le TYL, Raguram K, Hudson JE, et al. Platelet-derived growth factor-AB improves scar mechanics and vascularity after myocardial infarction. *Sci Transl Med.* 2020; 12.
70. Sutton MG, Sharpe N. Left ventricular remodeling after myocardial infarction: pathophysiology and therapy. *Circulation.* 2000; 101:2981-8.
71. Fan D, Takawale A, Lee J, Kassiri Z. Cardiac fibroblasts, fibrosis and extracellular matrix remodeling in heart disease. *Fibrogenesis Tissue Repair.* 2012; 5:15.
72. Maruhashi T, Kii I, Saito M, Kudo A. Interaction between periostin and BMP-1 promotes proteolytic activation of lysyl oxidase. *J Biol Chem.* 2010; 285:13294-303.
73. Trackman PC. Diverse biological functions of extracellular collagen processing enzymes. *J Cell Biochem.* 2005; 96:927-37.
74. Lopez B, Querejeta R, Gonzalez A, Beaumont J, Larman M, Diez J. Impact of treatment on myocardial lysyl oxidase expression and collagen cross-linking in patients with heart failure. *Hypertension.* 2009; 53:236-42.
75. Thompson NL, Flanders KC, Smith JM, Ellingsworth LR, Roberts AB, Sporn MB. Expression of transforming growth factor-beta 1 in specific cells and tissues of adult and neonatal mice. *J Cell Biol.* 1989; 108:661-9.
76. Heine U, Munoz EF, Flanders KC, Ellingsworth LR, Lam HY, Thompson NL, et al. Role of transforming growth factor-beta in the development of the mouse embryo. *J Cell Biol.* 1987; 105:2861-76.
77. Frangogiannis NG. The role of transforming growth factor (TGF)-beta in the infarcted myocardium. *J Thorac Dis.* 2017; 9:S52-S63.
78. Kallander LS, Washburn D, Hilfiker MA, Eidam HS, Lawhorn BG, Prendergast J, et al. Reverse Hydroxamate Inhibitors of Bone Morphogenetic Protein 1. *ACS Med Chem Lett.* 2018; 9:736-40.
79. Hanna A, Frangogiannis NG. The Role of the TGF-beta Superfamily in Myocardial Infarction. *Front Cardiovasc Med.* 2019; 6:140.
80. Dobaczewski M, Bujak M, Li N, Gonzalez-Quesada C, Mendoza LH, Wang XF, et al. Smad3 signaling critically regulates fibroblast phenotype and function in healing myocardial infarction. *Circ Res.* 2010; 107:418-28.
81. Kong P, Christia P, Frangogiannis NG. The pathogenesis of cardiac fibrosis. *Cell Mol Life Sci.* 2014; 71:549-74.
82. Uzel MI, Scott IC, Babakhanlou-Chase H, Palamakumbura AH, Pappano WN, Hong HH, et al. Multiple bone morphogenetic protein 1-related mammalian metalloproteinases process pro-lysyl oxidase at the correct physiological site and control lysyl oxidase activation in mouse embryo fibroblast cultures. *J Biol Chem.* 2001; 276:22537-43.
83. Butterfield RJ. Congenital Muscular Dystrophy and Congenital Myopathy. *Continuum (Minneapolis Minn).* 2019; 25:1640-61.
84. Mann CJ, Perdiguero E, Kharraz Y, Aguilar S, Pessina P, Serrano AL, et al. Aberrant repair and fibrosis development in skeletal muscle. *Skelet Muscle.* 2011; 1:21.
85. Nguyen Q, Lim KRQ, Yokota T. Current understanding and treatment of cardiac and skeletal muscle pathology in laminin-alpha2 chain-deficient congenital muscular dystrophy. *Appl Clin Genet.* 2019; 12:113-30.
86. Zambon AA, Muntoni F. Congenital muscular dystrophies: What is new? *Neuromuscul Disord.* 2021.
87. Yurchenco PD, McKee KK, Reinhard JR, Ruegg MA. Laminin-deficient muscular dystrophy: Molecular pathogenesis and structural repair strategies. *Matrix Biol.* 2018; 71-72:174-87.

88. Wilson SE, Marino GK, Torricelli AAM, Medeiros CS. Injury and defective regeneration of the epithelial basement membrane in corneal fibrosis: A paradigm for fibrosis in other organs? *Matrix Biol.* 2017; 64:17-26.
89. Accorsi A, Cramer ML, Girgenrath M. Fibrogenesis in LAMA2-Related Muscular Dystrophy Is a Central Tenet of Disease Etiology. *Front Mol Neurosci.* 2020; 13:3.
90. Taniguchi M, Kurahashi H, Noguchi S, Sese J, Okinaga T, Tsukahara T, et al. Expression profiling of muscles from Fukuyama-type congenital muscular dystrophy and laminin-alpha 2 deficient congenital muscular dystrophy; is congenital muscular dystrophy a primary fibrotic disease? *Biochem Biophys Res Commun.* 2006; 342:489-502.
91. Mehuron T, Kumar A, Duarte L, Yamauchi J, Accorsi A, Girgenrath M. Dysregulation of matricellular proteins is an early signature of pathology in laminin-deficient muscular dystrophy. *Skelet Muscle.* 2014; 4:14.
92. Mamuya FA, Duncan MK. α V integrins and TGF- β -induced EMT: a circle of regulation. *J Cell Mol Med.* 2012; 16:445-55.
93. Cencetti F, Bernacchioni C, Nincheri P, Donati C, Bruni P. Transforming growth factor- β 1 induces transdifferentiation of myoblasts into myofibroblasts via up-regulation of sphingosine kinase-1/S1P3 axis. *Mol Biol Cell.* 2010; 21:1111-24.
94. Contreras O, Rebolledo DL, Oyarzun JE, Olguin HC, Brandan E. Connective tissue cells expressing fibro/adipogenic progenitor markers increase under chronic damage: relevance in fibroblast-myofibroblast differentiation and skeletal muscle fibrosis. *Cell Tissue Res.* 2016; 364:647-60.
95. Elbaz M, Yanay N, Aga-Mizrachi S, Brunschwig Z, Kassis I, Ettinger K, et al. Losartan, a therapeutic candidate in congenital muscular dystrophy: studies in the dy(2J) /dy(2J) mouse. *Ann Neurol.* 2012; 71:699-708.
96. Meinen S, Lin S, Ruegg MA. Angiotensin II type 1 receptor antagonists alleviate muscle pathology in the mouse model for laminin-alpha2-deficient congenital muscular dystrophy (MDC1A). *Skelet Muscle.* 2012; 2:18.
97. Accorsi A, Kumar A, Rhee Y, Miller A, Girgenrath M. IGF-1/GH axis enhances losartan treatment in Lama2-related muscular dystrophy. *Hum Mol Genet.* 2016; 25:4624-34.
98. Barraza-Flores P, Hermann HJ, Bates CR, Allen TG, Grunert TT, Burkin DJ. Human laminin-111 and laminin-211 protein therapy prevents muscle disease progression in an immunodeficient mouse model of LAMA2-CMD. *Skelet Muscle.* 2020; 10:18.
99. Kuang W, Xu H, Vachon PH, Liu L, Loechel F, Wewer UM, et al. Merosin-deficient congenital muscular dystrophy. Partial genetic correction in two mouse models. *J Clin Invest.* 1998; 102:844-52.
100. Willmann R, Gordish-Dressman H, Meinen S, Ruegg MA, Yu Q, Nagaraju K, et al. Improving Reproducibility of Phenotypic Assessments in the Dy^W Mouse Model of Laminin-alpha2 Related Congenital Muscular Dystrophy. *J Neuromuscul Dis.* 2017; 4:115-26.